The interest in phenols and anthocyanins has increased due to their antioxidant properties and to their potential usage as dietary antioxidants in human nutrition. Total phenols and anthocyanin content, composition and stability in berry extracts of blackcurrant interspecific hybrids, and antioxidative activity of extracts was evaluated. Berries of interspecific hybrids accumulated 530 to 614 mg 100 g–1 FW of total phenolic compounds, while 621 mg 100 g–1 FW of phenolics was established in berries of control Ribes nigrum cultivar ‘Ben Tirran’. ‘Ben Tirran’ berries accumulated 444 mg 100 g–1 FW of anthocyanins and higher amount was identified in berries of interspecific hybrids No. 11–13 (R. nigrum × R. per- treum) and No. 57 (R. nigrum × R. aureum), 522 and 498 mg 100 g–1 FW respectively. Berry extracts of hybrid No. 11–13 distinguished by the highest antioxidative activity (80%) and it was higher than antioxidative activity of ‘Ben Tirran’ (70%). Antioxidative activity of all tested berry extracts (70–80%) was twice higher compared to synthetic antioxidant BHT (39%). However correlation between phenolics or total anthocyanin content and antioxidative activity degree was not established. Amount of cyanidin 3-O-glucoside and cyanidin 3-O-rutinoside in berries of interspecific hybrids No. 11–13 distinguished by the highest antioxidative activity (80%) and it was higher than antioxidative activity of ‘Ben Tirran’ (70%). Antioxidative activity of all tested berry extracts (70–80%) was twice higher compared to synthetic antioxidant BHT (39%). However correlation between phenolics or total anthocyanin content and antioxidative activity degree was not established. Amount of cyanidin 3-O-glucoside and cyanidin 3-O-rutinoside in berries of interspecific hybrids No. 57, No. 11–13 and No. 8 (R. nigrum × R. americanum) was higher than in berries of ‘Ben Tirran’. It was established that cyanidins are more stable anthocyanins in all studied temperature and irradiation conditions. Therefore interspecific hybrids No. 57 and No. 11–13 were the most agronomically valuable hybrids.

Key words: anthocyanins, antioxidative activity, interspecific hybrids, phenolics, stability

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INTRODUCTION

Search of bioactive plant materials important to human health is one of problems in biomedical research (Pourmorad et al., 2006; Pascual-Teresa & Sanches-Ballesta, 2008; Borges et al., 2010; Bunca et al., 2011). Bioactive substances like anthocyanins, other phenolic compounds, including tannins, proanthocyanidins and phenolic acids derived from berries have received considerable interest (Moyer et al., 2002; Horbowicz et al., 2008; Arnnok et al., 2012; Tsuda, 2012). The interest in anthocyanins has increased due to their antioxidant properties and to their potential usage as dietary antioxidants in human nutrition (Pascual-Teresa & Sanches-Ballesta, 2008; Lugasi et al., 2011). The most important role of anthocyanins is the antioxidant activity, which includes protection against DNA damage, free radical scavenging, coursing depletion of immune system antioxidants, change in gene expression and induce formation the abnormal proteins, involved in many health disorders (Pourmorad et al., 2006; Oancea & Opren, 2011). It is known, that these compounds are being able to capture reactive oxygen species, such as superoxide radical — O2–, hydroxyl radical – HO –, hydrogen peroxide — H2O2 or singlet oxygen –1O2, can delay the initiation or propagation of oxidative chain reactions, inhibits the oxidation (Javanmardi et al., 2003; Denex et al., 2012). Numerous publications are presented in which the antioxidative activity of anthocyanins extracted from berries or fruits is indicated (Pantelidis et al., 2007; Horbowicz et al., 2008; Djordjević et al., 2010). To date the potential effect of anthocyanins, other phenolic compounds in reducing the incidence of cardiovascular disease prevention, cancer, diabetes, hyperlipidemias, possessing anti-microbial, anti-inflammatory and anti-carcinogenic activity, obesity control, and other chronic diseases are indicated (Pascual-Teresa & Sanches-Ballesta, 2008; Liegiūtė et al., 2009; Liobikas et al., 2009; Shipp & Abdel-Âal, 2010; Oancea & Opren, 2011; Tsuda, 2012).

More than 635 natural anthocyanins were identified, they are constituted from anthocyanidins which are differently glycosylated and acylated (He & Giusti, 2010), although the relationship between their function and structure are disputable and remains under consideration. Anthocyanins lacking the O-diphenyl structure in the B ring (malvidin, pelargonidin, petunidind and penonidin) have a lower DPPH radical scavenging efficiency compared to cyanidin and delphinidin. Position of hydroxyl and methylation in the B ring affects the stability, antioxidative activity, anti-carcinogenic activity, obesity control, and other chronic diseases are indicated (Pascual-Teresa & Sanches-Ballesta, 2008; Liegiūtė et al., 2009; Liobikas et al., 2009; Shipp & Abdel-Âal, 2010). The composition of anthocyanins in different plant species berries or fruits is not enough investigated and we have knowledge gaps whether composition of anthocyanins and/or which alone anthocyanin have the most significant impact in their antioxidative capacity.
Content, composition and antioxidant activity of anthocyanins and other phenolic compounds in berries depends not only on genotype, but also on environmental factors (Blando et al., 2004; Pantelidis et al., 2007; Horbowicz et al., 2008). Therefore anthocyanin content, composition and activity in plants, grown in different countries should be studied. The growing interest of antioxidant influence on human health (Guerrero et al., 2010) has triggered the identification of new producers of phenolics compounds and anthocyanins, which distinguishes in their antioxidative activity. The berries of Ribes genus are considered to be rich in anthocyanins and total phenolics in various countries (Slimestad & Solheim, 2002; Rubinskienė et al., 2005; Horbowicz et al., 2008), breeding programs were aimed to increase berry nutritional value.

The aim of our study was to analyse total phenols and to evaluate anthocyanin content, composition and stability in berry extracts of blackcurrant interspecific hybrids, to evaluate antioxidative activity of extracts and to identify perspective hybrids — a new sources of anthocyanins and phenols with high antioxidative capacity.

**MATERIALS AND METHODS**

**Plant material.** Anthocyanins, total phenols and antioxidative activity of berry extracts of blackcurrant (Ribes nigrum) ‘Ben Tirran’ and complex interspecific hybrids of blackcurrant with american currant (R. americanum), golden currant (R. aureum), gooseberry (R. uva-crispa) and red currant (R. petraeum). No. 6 (R. nigrum × R. americanum) × (R. nigrum × R. aureum) × R. uva-crispa, No. 8 (R. nigrum × R. americanum) × (R. nigrum × R. americanum), No. 11–13 R. nigrum × R. petraeum, No. 57 R. nigrum × R. aureum were studied. Berries were collected at technical maturity phase, immediately frozen and stored at −70°C until analysis.

**Extraction of phenolic compounds and anthocyanins.** Anthocyanins and other phenolics were extracted from frozen berries ground to a fine powder using 90% aqueous methanol, acidified with HCl to 0.1 N at ratio 1:20 g/ml and stored for 16 h at 4°C in the dark (Anisimovienė et al., 2009; Arrnok et al., 2012). Reagents were purchased from Sigma-Aldrich. The material was shaken twice for 30 min at 4°C in the dark during the extraction procedure. The extract was filtrated through membrane filters (0.2-μm pore diameter, Whatman) residues were washed until solvent became colourless.

The obtained methanol extracts were used for determination of anthocyanins and total phenolics and for establishment of antioxidative activity. Later extracts were vacuum dried, using an IKA RV-10 water pump (Germany) at 40°C. Samples were stored at −70°C until analysis of anthocyanin composition using high performance liquid chromatography method (HPLC) and for study of stability of antioxidative activity.

**Determination of total anthocyanins.** The anthocyanins content was determined using a spectrophotometric differential pH method (Wrolstad et al., 2005; Horbowicz et al., 2008). Absorbance of the extracts was measured at 520 and 700 nm, using a UV-VIS spectrophotometer (Specord 210 PLUS, Analytik Jena AG, Jena, Germany). Buffers: pH 1.0 (hydrochloric acid–potassium chloride 0.025 M) and pH 4.5 (hydrochloric acid–sodium acetate 0.4 M). Exposition time was 30 min, at 25°C in dark. Anthocyanins content was calculated using a molar extinction coefficient of cyanidin-3-glucoside chloride (ε3g). Results were expressed as cyanidin-3-glucoside equivalents mg 100 g−1 fresh weight (FW). Reagents were purchased from Sigma-Aldrich.

**Determination of phenolic compounds.** Total phenolics content in crude methanol extracts was determined by Folin-Ciocalteu method Slinkard and Sigleton (1977) as referred by (Pantelidis et al., 2007). Folin-Ciocalteu’s reagent (Fluka) was diluted with distilled water (1/10, v/v) and 7.5% aqueous Na2CO3 (both from Sigma-Aldrich) were used for measurement. Exposition time was 10 min, at 25°C in dark. Absorbance of samples was measured at 765 nm, using spectrophotometer (Specord 210 PLUS, Germany). The gallic acid (Sigma-Aldrich) was used as a standard. Phenolic compound content was expressed as mg 100 g−1 FW.

**Determination of antioxidant activity.** The antioxidant activity (capacity) of methanol extracts derived from various berries was determined spectrophotometrically according a free radical scavenging activity, using the 2,2-diphenyl-1-picrylhydrazyl (DPPH) method (Anisimovienė et al., 2009; Viskelis et al., 2010; Bunea et al., 2011; Hassanpour et al., 2011). DPPH was purchased from Sigma-Aldrich. Fifty microliters of methanol extracts derived from various berries (concentrations 13–16 mg mL−1) were mixed with 1.0 mL−1 of 6 × 10−3 mol L−1 of freshly prepared DPPH. Mixtures were kept in dark at 25°C for 30 min. The absorbance was measured at 515 nm, using spectrophotometer (Specord 210 PLUS, Analytik Jena AG, Jena, Germany). The antioxidative activity — inhibition of DPPH radical (%) was evaluated according the absorbance of the blank solution and absorbance value of sample solution and expressed in percents (%). The antioxidative activity (%) of tested materials was compared to antioxidant activity of synthetic inhibitor 2,6-di-tret-butyl-4-methyl-phenol (BHT) (Sigma-Aldrich).

**Determination of anthocyanins composition.** The composition of anthocyanins in prepared extracts was investigated using HPLC (Durst & Wrolstad, 2001; Libobikas et al., 2009). Anthocyanin analysis was performed using Agilent 1200 HPLC system with DAD detector (Agilent, Germany); reverse phase XBridge Shield RP18 (Waters, UK) analytical column (3.0 × 150 mm, particle size 3.5 μm) was used. The mobile phase consisted of 10% acetic acid and 1% phosphoric acid (solvent A) and 100% acetonitrile (solvent B), HPLC ultra gradient grade reagents were purchased from Sigma-Aldrich. The elution conditions were as follows: isocratic elution 0% B, 0–12 min; linear gradient from 0% B to 7% B, 12–15 min; to 45% B, 17 min; to 100% B, 20 min, flow rate 0.7 ml min−1. Detector wavelength of 520 nm was used. Anthocyanin standards: cyanidin 3-O-glucoside (c3g), cyanidin 3-O-rutinoside (c3r), delphinidin 3-O-rutinoside (d3r), delphinidin 3-O-glucoside (d3g), pelargonidin 3-O-glucoside (p3g), peonidin 3-O-rutinoside (p3r) and malvidin 3-O-glucoside (m3g) (Extrasynthese, France; Polyphenols Laboratories AS, Norway) were used for identification and quantification of individual anthocyanins.

**Evaluation of anthocyanin stability in berry extracts.** Stability of anthocyanins in berry extracts of interspecific hybrid No. 57 was evaluated at different irradiances and temperature conditions. Storing conditions: +23 ± 2°C with 50–150 μmol m−2 s−1 photosynthetic photon flux density; +23 ± 2°C in dark and +4 ± 1°C in dark. Anthocyanin content and composition was evaluated after 2, 7, 14, 28, 56 and 84 days of storage using HPLC.

**Statistical analysis.** Average data and ± S.E. are presented for 5 repetitions. The data were computed using...
the Microsoft Excel Descriptive Statistics program at the 95% confidence level.

RESULTS AND DISCUSSION

Content of phenolics in berries of interspecific blackcurrant hybrids

The highest amount of total phenolic compounds (above 600 mg 100 g⁻¹ FW) was established in berries of ‘Ben Tirran’ and hybrid No. 11–13 (R. nigrum × R. petraeum) (Fig. 1). Berries of interspecific Ribe hybrids accumulated less phenolics, than control variety ‘Ben Tirran’. According other studies, higher phenolics content is characteristic to blackcurrant compared to gooseberries and redcurrants (Moyer et al., 2002; Horbowicz et al., 2008; Pinto et al., 2010). Therefore lower amount of phenolics in interspecific hybrids was due blackcurrant hybridisation with these species.

Content of anthocyanins in berries of interspecific blackcurrant hybrids

The blackcurrants are recognized as a good source of anthocyanins and are valued for their health benefits. The anthocyanin content established in blackcurrant berries in Lithuania and countries (Horbowicz et al., 2008; Nour et al., 2011). Our results show that blackcurrant berries in Lithuania are rich sources of anthocyanins.

Antioxidative activity of berry extracts of interspecific blackcurrant hybrids

One of the most important roles of anthocyanins is their antioxidative activity — capability to scavenge the free radicals (Onacea & Optean, 2011). Antioxidative activity showed no significant variations between tested blackcurrants and it ranged from 69% to 80% (Fig. 3). Besides that, antioxidative activity of all tested extracts was twice higher compared to synthetic antioxidant butylated hydroxytoluene (BHT). Also, antioxidative activity of Ribe berry extracts was higher compared to blueberries, elderberries (Anisimoviene et al., 2009).

The highest antioxidative activity was identified in berry extracts of hybrid No. 11–13 (R. nigrum × R. petraeum) and No. 6 ((R. nigrum × R. americanum) × (R. nigrum × R. aureum × R. uva-crispa)) and it was higher than antioxidative activity of a control blackcurrant cultivar ‘Ben Tirran’.
between different anthocyanin compounds. Berry extracts may be related to composition and ratio of anthocyanins and phenolics (Shipp, Abdel-Ala, 2010). Though antioxidative activity of berry extracts depends not only on anthocyanins but on stilbenes, ellagitanins, tanins, proanthocyanidins, or cyanidin 3-O-rutinoside, cyanidin 3-O-galacoside, malvidin 3-O-glucoside, x — unidentified anthocyanins.

Our results support the theory, that antioxidant effect of berry extracts depends not only on anthocyanins but on stilbenes, ellagitanins, tanins, proanthocyanidins, phenolic acids also (Maatta et al., 2001; Heinonen, 2007; Shipp, Abdel-Ala, 2010). Though antioxidative activity of berry extracts may be related to composition and ratio between different anthocyanins compounds.

Table 1. Anthocyanin composition in berry extracts of 'Ben Tirran' and interspecific Ribes hybrids.

<table>
<thead>
<tr>
<th>Cultivar, hybrid</th>
<th>d3g</th>
<th>d3r</th>
<th>c3g</th>
<th>c3r</th>
<th>pel3g</th>
<th>peo3r</th>
<th>peo3g</th>
<th>m3g</th>
<th>x</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. ('Ben Tirran')</td>
<td>78.56</td>
<td>±2.55</td>
<td>200.39</td>
<td>±6.80</td>
<td>14.57</td>
<td>±0.52</td>
<td>120.90</td>
<td>±4.75</td>
<td>0.64</td>
</tr>
<tr>
<td>2.</td>
<td>±6.36</td>
<td>±26.88</td>
<td>±5.51</td>
<td>±17.17</td>
<td>±0.23</td>
<td>–</td>
<td>–</td>
<td>±2.03</td>
<td>±1.37</td>
</tr>
<tr>
<td>3.</td>
<td>116.03</td>
<td>±7.94</td>
<td>120.99</td>
<td>±14.33</td>
<td>49.69</td>
<td>±2.24</td>
<td>110.44</td>
<td>±11.40</td>
<td>5.04</td>
</tr>
<tr>
<td>4.</td>
<td>70.71</td>
<td>±6.24</td>
<td>262.53</td>
<td>±7.92</td>
<td>25.03</td>
<td>±1.77</td>
<td>146.04</td>
<td>±8.06</td>
<td>4.26</td>
</tr>
<tr>
<td>5.</td>
<td>±3.06</td>
<td>±58.41</td>
<td>±29.02</td>
<td>±32.05</td>
<td>±2.07</td>
<td>–</td>
<td>–</td>
<td>±1.39</td>
<td>±0.31</td>
</tr>
</tbody>
</table>

Mean ± S.E. are presented, n=5.

Anthocyanin composition in berries of interspecific blackcurrant hybrids

Anthocyanin content and amount in berries of blackcurrant 'Ben Tirran' and interspecific asymmetric hybrids of blackcurrant with currant species depending to other sections was evaluated using HPLC (Table 1). 6–7 Anthocyanins were identified in all the extracts; however 2–5 anthocyanins remained unidentified in each sample. Berry extracts of 'Ben Tirran' and hybrid No. 11–13 (R. nigrum × R. petraeum) had 7 anthocyanins, while 6 anthocyanins were identified in other hybrids berry extracts. Delphinidin 3-O rutinoside was dominant anthocyanin in all studied blackcurrants; it constituted 28–50 percent of total anthocyanin content. Amount of d3g was higher in berries of hybrids No. 8 ((R. nigrum × R. americanum) × (R. nigrum × R. americanum)) and No. 57 (R. nigrum × R. aureum) than in control cultivar 'Ben Tirran'.
Amount of cyanidin-3-O glucoside, which is stable anthocyanin, was 1.7–3.3 times higher in berries of all hybrids than in berries of blackcurrant ‘Ben Tirran’. C3g content in berries of hybrids No. 57 and No. 11–13 was significantly larger than in berries of ‘Ben Tirran’. Similar C3r amount was established in berry extracts of hybrids No. 6 ([R. nigra × R. americanum] × [R. nigra × R. arunium] × R. urs-crispa), No. 8 and blackcurrant ‘Ben Tirran’, but was higher in No. 11–13 and No. 57. Malvidin-3-O amount ranged from 1.39 to 5.25 mg 100 g–1 in all studied berries, while peonidin-3-O glucoside was found in ‘Ben Tirran’ and peonidin-3-O rutinoside in hybrid No. 11–13 berries only. The largest quantity of unidentified anthocyanins was established in berries of ‘Ben Tirran’ 24.92 mg 100 g–1 FW (5.6 percent of total anthocyanins), while in berries of interspecific hybrids amount of unidentified anthocyanins was lower 3.77–16.22 mg 100 g–1 FW (1.0–3.8 percent).

Evaluation of anthocyanin stability in berry extracts

In order to select perspective interspecific currant hybrids with high amount of stable anthocyanins it is important to evaluate stability of individual anthocyanins. Stability of anthocyanins in berry extracts of interspecific hybrid No. 57 ([R. nigra × R. arunium] during storage at different lighting and temperature conditions was evaluated (Fig. 4).

Four anthocyanins (d3g, d3r, c3r, c3g) were dominant in berries of interspecific hybrid No.57, they constituted 98.4% of all anthocyanins. After storage in room temperature in light for 14 days anthocyanin amount decreased, and only 19–23 % of delphinidins remained, while cyanidins were more stable (41–44% remained). Only 3–4% of delphinidins and 18–21% of cyanidins remained non degraded after storage for 28 days, and almost all anthocyanins were degraded after 56 days. Degradation of anthocyanins in the same extracts, stored at room temperature in dark was slower. More than 69% of cyanidins and 49–53% of delphinidins remained stable after storage for 28 days, and 40–44% of cyanidins and 15–19% of delphinidins were non degraded after storage for 84 days (Fig. 4). Eighty six to ninety two percent of both delphinidins and cyanidins were more stable after storage in dark and cold (+4oC) conditions after 28 days, and after 84 days in these conditions 72 percent of cyanidins and 59% of delphinidins were non degraded. Other studies present similar anthocyanin stability data (Hellström et al., 2013), but they studied total anthocyanin content only, and our study reveals stability of individual anthocyanin in berry extract.

Data on phenols, anthocyanin quantity and composition show that two interspecific hybrids (No. 11–13 and No.57) equal and exceed ‘Ben Tirran’. Thus, the new interspecific hybrids may be characterized as possessing a high health benefits.

CONCLUSIONS

Amount of total phenolic compounds in berries of interspecific hybrids ranged from 530 to 614 mg 100 g–1 FW, Ribes nigra ‘Ben Tirran’ berries accumulated 621 mg 100 g–1 FW. The highest amount of anthocyanins was established in berries of hybrids No. 11–13 ([R. nigra × R. petraeum] × [R. nigra × R. arunium]) and after 84 days in these conditions 72 percent of cyanidins and 49–53% of delphinidins remained stable after storage. More than 69% of cyanidins and 49–53% of delphinidins remained stable after storage for 28 days, and 40–44% of cyanidins and 15–19% of delphinidins were non degraded after storage for 56 days. Degradation of anthocyanins in the same extracts, stored at room temperature in dark was slower. More than 69% of cyanidins and 49–53% of delphinidins remained stable after storage for 28 days, and 40–44% of cyanidins and 15–19% of delphinidins were non degraded after storage for 84 days (Fig. 4). Eighty six to ninety two percent of both delphinidins and cyanidins were more stable after storage in dark and cold (+4oC) conditions after 28 days, and after 84 days in these conditions 72 percent of cyanidins and 59% of delphinidins were non degraded. Other studies present similar anthocyanin stability data (Hellström et al., 2013), but they studied total anthocyanin content only, and our study reveals stability of individual anthocyanin in berry extract.

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